

DINOSTEROL SIDE CHAIN BIOSYNTHESIS IN A MARINE DINOFLAGELLATE, *CRYPTHOCODINIUM COHNII*

NANCY W. WITHERS,* ROBERT C. TUTTLE, L. JOHN GOAD and TREVOR W. GOODWIN

Department of Biochemistry, University of Liverpool, P.O. Box 147, Liverpool L69 3BX, U.K.

(Received 12 June 1978)

Key Word Index—*Cryptocodinium cohnii*; dinoflagellate; dinosterol; methionine-[CD₃]; side chain biosynthesis.

Abstract—The heterotrophic dinoflagellate, *Cryptocodinium cohnii*, cultured in a nutrient medium containing methionine-[CD₃] incorporated deuterium into the newly synthesized 4 α -monomethyl compound dinosterol (4 α ,23,24-trimethylcholestan-22-en-3 β -ol). The MS fragmentation pattern indicated that the C-23 methyl group contained three deuterium atoms and was introduced intact by transmethylation from methionine. The C-24 methyl group contained only two deuterium atoms which is consistent with the production of a 24-methylenesterol intermediate which is subsequently reduced to give the 24-methyl side chain. Mechanisms are proposed to account for the production of the dinosterol side chain.

INTRODUCTION

Studies on the mechanisms of phytosterol side chain alkylation at C-24 have been facilitated by the use of the deuterium labelled methyl group donor, methionine-[CD₃] added to the growth media and subsequent mass spectroscopic examination of newly synthesized sterols to determine their deuterium content [1, 2]. In this way the formation of the C-24 ethyl or methyl groups of the sterols produced by chrysophyte [3], bacillariophyte [4] and chlorophyte [5, 6] algae have been studied.

The structural elucidation of dinosterol (1), isolated from a dinoflagellate *Gonyaulax tamarensis*, as (22*E*, 24*R*)-4 α ,23,24-trimethyl-5 α -cholestan-22-en-3 β -ol [7, 8], revealed that this alga produced a C-23 methylated sterol. The only other examples of C-23 methylated sterols are the marine invertebrate sterols gorgosterol and acanthasterol isolated from soft corals [9] and an echinoderm [10], respectively. Our demonstration of the production of dinosterol (1) in *Cryptocodinium cohnii* [11], a heterotrophic dinoflagellate which can be cultured on a defined

synthetic medium [12], has enabled us to study the mechanism of alkylation at C-23 and C-24 in this unique phytosterol.

RESULTS AND DISCUSSION

Examination by MS of the sterols isolated from *C. cohnii* cultured in the presence of methionine-[CD₃] revealed that over 60% of the newly synthesized 4 α -

Table 1. MS fragmentation ions observed for dinosterol (1) isolated from *Cryptocodinium cohnii* grown in the presence of methionine-[CD₃]

Fragmentation*	M ⁺	Parent molecular ion species		
		M ⁺ + 2D	M ⁺ + 3D	M ⁺ + 5D
M ⁺	428	430	431	433
M ⁺ - a(43)	385	387	388	390
M ⁺ - a-H ₂ O	367	369	370	372
M ⁺ - b(71)	357	357	360	360
M ⁺ - b-H ₂ O	339	339	342	342
M ⁺ - c(111)-H	316	316	316	316
M ⁺ - d-2H	287	287	287	287

* See Fig. 1. for manner of fragmentation.

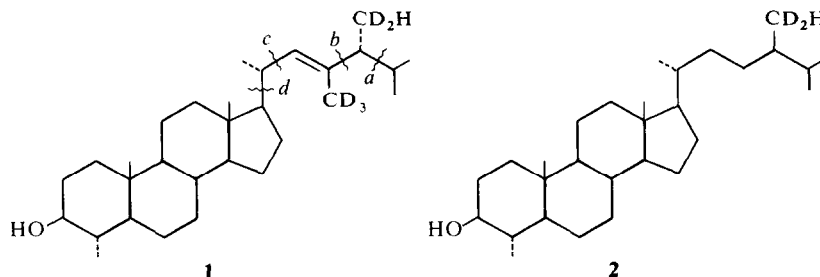


Fig. 1. Dinosterol (1) and 4 α ,24-dimethylcholestan-3 β -ol (2) showing deuterium labelling patterns obtained from culture of *C. cohnii* in the presence of methionine-[CD₃]. The MS fragmentation of the side chain of 1 is indicated (see Table 1).

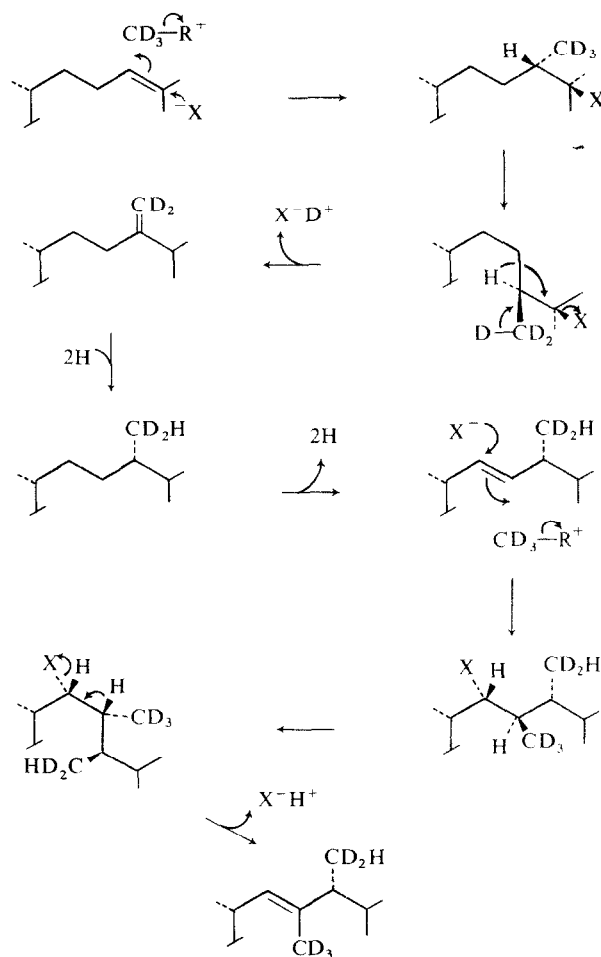


Fig. 2. Postulated mechanisms for biosynthetic side chain alkylations at C-23 and C-24 in dinosterol (1) production in *C. cohnii*.

monomethyl sterols contained deuterium atoms whereas no deuterium was incorporated into cholesterol or cholesta-5,7-dien-3 β -ol which are the major 4-demethylsterols in this dinoflagellate [11].

The MS of the 4-monomethyl compound dinosterol (1) showed it to contain molecular species possessing two, three and five deuterium atoms (Table 1). The fragmentations at *a* and *b* (Fig. 1) produced ions containing a maximum of five and three deuteriums, respectively (Table 1), thus revealing the presence of $-\text{CD}_2\text{H}$ at C-24. This is compatible with the production of a 24-methylene intermediate in the C-24 alkylation sequence which is subsequently reduced to produce the C-24 methyl group of dinosterol (1). This mechanism has been reported previously to operate in fungi [1], a chrysophyte alga [3] and a diatom [4]. The fragmentations at *b* and *c* or *d* gave ions containing three and no deuterium atoms respectively which therefore demonstrated that the C-23 methyl group was derived intact by transmethylation from the methionine donor with retention of all three deuterium atoms. A second 4 α -methyl-sterol from *C. cohnii*, which we have tentatively identified [11] 4 α ,24-dimethyl-cholestan-3 β -ol (2), showed MS molecular ions at *m/e* 416 and 418, the retention of two deuterium atoms being in accord with a 24-methylene

intermediate as concluded above for production of the C-24 methyl group of dinosterol (1).

A suggested biosynthetic scheme based upon the present results for the elaboration of the dinosterol (1) side chain is presented in Fig. 2. The alkylation steps involved in dinosterol (1) side chain biosynthesis may be relevant to considerations concerning the biosynthetic origins of the side chain of gorgosterol, (22*R*, 23*R*, 24*R*)-22,23-methylene-23,24-dimethylcholest-5-en-3 β -ol [9], which is found in gorgonian corals containing symbiotic zooxanthellae which are apparently dinoflagellates [13].

EXPERIMENTAL

Cryptocodinium cohnii (Woods Hole Strain d) was grown in 11. of modified MLH medium [12] with reduced betaine (0.15 g/l) and added methionine- $[\text{CD}_3]$ (1.5 g/l). Sterols were isolated and analysed as described previously [11]. MS were determined by direct probe at 70 eV in an AEI MS 12 instrument.

Acknowledgements—NWW gratefully acknowledges support of a Leverhulme Visiting Fellowship (1976–77). MS were determined by Mark Prescott and the work was financed by the SRC.

REFERENCES

1. Lederer, E. (1964) *Biochem. J.* **93**, 449.

2. Goad, L. J., Knapp, F. F., Lenton, J. R. and Goodwin, T. W. (1974) *Lipids* **9**, 582.
3. Smith, A. R. H., Goad, L. J., Goodwin, T. W. and Lederer, E. (1967) *Biochem. J.* **104**, 58C.
4. Rubinstein, I. and Goad, L. J. (1974) *Phytochemistry* **13**, 485.
5. Tomita, Y., Uomori, A. and Sakurai, E. (1971) *Phytochemistry* **10**, 573.
6. Goad, L. J., Lenton, J. R., Knapp, F. F. and Goodwin, T. W. (1972) *Biochem. J.* **129**, 219.
7. Shimizu, Y., Alam, M. and Kobayashi, A. (1976) *J. Am. Chem. Soc.* **98**, 1059.
8. Finer, J., Hirotsu, K. and Clardy, J. (1977) in *Marine Natural Products Chemistry* (Faulkner, D. J. and Fenical, W. H., eds.) p. 147. Plenum Press, New York.
9. Ling, N. C., Hale, R. L. and Djerassi, C. (1970) *J. Am. Chem. Soc.* **92**, 5281.
10. Sheikh, Y. M., Djerassi, C. and Tursch, B. M. (1971) *Chem. Commun.* 217.
11. Withers, N. W., Tuttle, R. C., Holz, G. G., Beach, D. H., Goad, L. J. and Goodwin, T. W. (1978) *Phytochemistry*, **17**, 1987.
12. Tuttle, R. C. and Loeblich, A. R., III (1975) *Phycologia* **14**, 1.
13. Jeffrey, S. W. and Haxo, F. T. (1968) *Biol. Bull.* **135**, 149.